



Tetranortriterpenoid derivatives from *Turraea parvifolia* (Meliaceae)

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Abstract

The methanol extract of the seeds of *Turraea parvifolia* has yielded seven novel triterpenoid derivatives: 12 α -acetoxyazadironolide, turrarparvin A (12 α -acetoxy-7 α ,23-dihydroxy-24,25,26,27-tetranor-3-oxoapotirucalla-1,14,20(22)-trien-21,23-olide), turrarparvin B (12 α -acetoxy-7 α ,21-dihydroxy-24,25,26,27-tetranor-3-oxoapotirucalla-1,14,20(22)-trien-23,21-olide), turrarparvin C (7 α ,12 α -diacetoxy-21-hydroxy-24,25,26,27-tetranor-3-oxoapotirucalla-1,14,20(22)-trien-23,21-olide), 11-*epi*-21-hydroxytoonacilide, 11-*epi*-23-hydroxytoonacilide and turrarparvin D (12 α -acetoxy-7 α -hydroxy-24,25,26,27-tetranor-3-oxoapotirucalla-1,14,20(22)-trien-21,23-lactam).

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Keywords: Meliaceae; *Turraea parvifolia*; 12 α -Acetoxyazadironolide; Turrarparvin A; Turrarparvin B; Turrarparvin C; 11-*epi*-21-Hydroxytoonacilide, 11-*epi*-23-Hydroxytoonacilide and turrarparvin D

1. Introduction

Turraea parvifolia Deff. (Meliaceae) is a small tree or shrub found in East Africa. It is usually found in open *Acacia-Commiphora* bush-land, especially around rock outcrops and on banks of seasonal watercourses. It is characterized by its smooth, dark grey stems, slender drooping branches and small white flowers (Blundell, 1987). It is known as “Chositim” by the Pokot tribe of Kenya, where it is used as an emetic.

The genus *Turraea* (Meliaceae) is placed in the tribe Turraeeae, subfamily Melioideae (Styles and White, 1991). The genus comprises some 60–70 species of shrubs and small trees widely distributed in eastern Africa and the islands of the Indian Ocean. *Turraea* species are widespread, but they are nowhere common, or at least are inconspicuous and, therefore, difficult to obtain (Bentley et al., 1995; Mulholland and Taylor, 1988). Seven species of the genus *Turraea* have been investigated chemically and a number of limonoids, protolimonoids and triterpenoids have been isolated (Adul et al., 1993; Akinniyi et al., 1986; Bentley et al., 1992, 1995; Chiplunkar et al., 1993; Fraser et al., 1994, 1995; Mulholland and Taylor, 1988; Mulholland et al., 1993, 1998, 1999; Rajab et al., 1988; Torto et al., 1995, 1996).

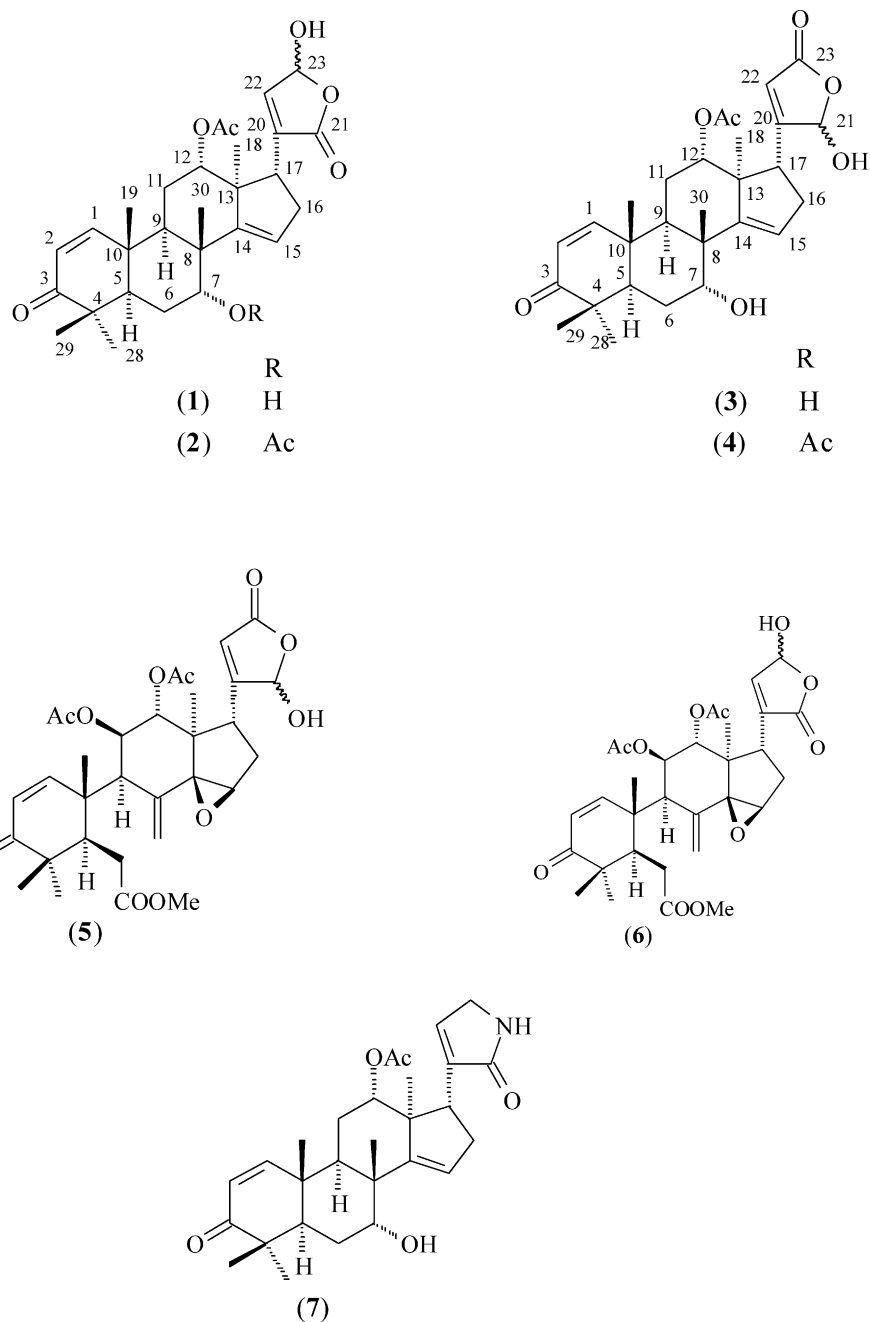
2. Results and discussion

Six novel triterpenoid derivatives and one novel tetranortriterpenoid lactam were isolated from the methanol extract of the seed of *Turraea parvifolia*. As six of the compounds isolated were hydroxybutenolides and these occur as epimeric mixtures of the interconverting α - and β -hydroxy compounds in solution, NMR spectra were complex and some pairing of some peaks occurred (Kraus and Grimminger, 1980; Siddiqui et al., 1999). The peak due to the major epimer is given in the text, with that due to the minor one in brackets.

High resolution mass spectrometry of compound **1**, turrarparvin A, showed no molecular ion peak, however, fragment ion peaks were observed at m/z 466.2367 [$M^+ - 18$], 424.2251 [$M^+ - 60$] and 406.2119 [$M^+ - 18 - 60$] due to the loss of water, acetic acid or both water and acetic acid. The molecular formula of $C_{28}H_{36}O_7$ was deduced from the mass spectrum in conjugation with the ^{13}C NMR spectrum. Rings A–D were intact, an α,β -unsaturated ketone in ring A was indicated by a pair of doublets at δ 7.01 ($J = 10.2$ Hz, H-1) and δ 5.81 ($J = 10.2$ Hz, H-2) and resonances at δ 157.45 (157.41), δ 125.60 and δ 204.98 (204.95) ascribed to C-1, C-2 and C-3 respectively. A Δ^{14} -double bond was indicated by resonances at δ 158.46 (158.80) and δ 121.75 (121.95) for C-14 and C-15 respectively, and a resonance ascribed to H-15 at δ 5.64.

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A hydroxy and an acetate group were present at C-7 α and C-12 α respectively. The H-7 resonance at δ 4.02 was assigned based on HMBC correlation with C-5 and a NOESY correlation with 3H-30, confirmed H-7 was β and thus, the hydroxy group was α . The H-12 resonance at δ 4.95 showed HMBC correlations with C-18 and C-17. The H-12 resonance showed a correlation with the 3H-30 resonance and the acetate methyl group proton resonance showed NOESY correlations with 3H-18 and H-9, confirming that the acetoxy group was in the α -orientation. The presence of a 23-hydroxy-21,23-butenolide side chain was indicated by a C-21 lactone carbonyl carbon resonance at δ 171.38 (171.33), a C-20,C-22 double

bond indicated by resonances at δ 137.03 (137.94) and δ 144.92 (145.57). The H-22 (δ 6.95) resonance was seen to be coupled to H-23 in the COSY spectrum and C-23 occurred at δ 96.40 (96.12). The relative stereochemistry at C-5, C-9 and C-17 was confirmed for all compounds isolated by means of the NOESY spectrum. The H-9 α resonance was seen to correlate with 3H-18 and 3H-28 (which are α) and H-5 also showed a correlation with the 3H-28 resonance. The H-17 resonance showed a NOESY correlation with H-12 β confirming the β -orientation of H-17. All ^1H and ^{13}C NMR resonances could be assigned using COSY, HMBC and NOESY spectra and are given in [Tables 1 and 2](#).

Compound **2** (C₃₀H₃₈O₈) was identified as 12 α -acetoxyazadirone, **2**. This compound is the 7-acetyl derivative of turrarparvin A and this was confirmed by acetylation of turrarparvin A which gave 12 α -acetoxyazadirone. This compound is a derivative of azadirone previously isolated from *Azadirachta indica* (Siddiqui et al., 1999).

Compound **3** (C₂₈H₃₆O₇), turrarparvin B, differed from **1** in the structure of the side-chain. Instead of a 23-hydroxy-21,23-butenolide ring, a 21-hydroxy-23,21-butenolide ring was present in **3**. This was confirmed by HMBC correlations. Analogously, compound **4**, turrarparvin C, (C₃₀H₃₈O₈) differs from compound **2** in having the 23-hydroxy-21,23-butenolide ring instead of the 21-hydroxy-23,21-butenolide ring.

The NMR spectra of compound **5**, 11-*epi*-21-hydroxytoonacilide, (C₃₁H₃₈O₁₁), showed that it was a ring B opened limonoid. Two H-30 resonances occurred as broad singlets at δ 5.34 and δ 5.27 and C-8 and C-30

occurred at δ 135.68 (135.49) and δ 122.29 (121.90) respectively. A carbomethoxy group was present at C-7 and the NMR spectrum showed C-7 occurring at δ 174.14 and the methoxy group proton resonance at δ 3.66 as is usual in ring-B opened limonoids (Kraus and Grimmer, 1980). An α,β -unsaturated ketone in ring A was indicated by a pair of doublets at δ 7.36 ($J=10.6$, H-1) and δ 6.12 ($J=10.6$, H-2) and resonances at δ 151.96 (152.07), δ 125.87 and δ 204.30 (204.19) ascribed to C-1, C-2 and C-3 respectively. No Δ^{14} -double bond was present, but a 14,15-epoxide was indicated by resonances at δ 71.30 (70.61) and δ 59.47 (58.97) and a resonance ascribed to H-15 at δ 3.92. The 14,15-epoxide was assigned a β -orientation due to a NOESY correlation between H-15 and H-30. Proton resonances ascribable to two acetoxy groups were present at δ 1.94 and δ 1.91 in the ¹H NMR spectrum. These were placed at C-11 and C-12 due to the presence of a H-9 (δ 2.96), H-11 (δ 5.54), H-12 (δ 5.65) coupled system seen in the

Table 1

NMR spectral data for compounds **1–7** (400 MHz, CDCl₃) (J , in Hz, given in parentheses)

¹ H	1	2	3	4	5	6	7
1	7.01 <i>d</i> (10.2)	7.02 <i>d</i> (10.3)	7.01 <i>d</i> (10.3)	7.01 <i>d</i> (10.3)	7.36 <i>d</i> (10.6)	7.37 <i>d</i> (10.5)	7.00 <i>d</i> (10.2)
2	5.81 <i>d</i> (10.2)	5.81 <i>d</i> (10.1)	5.80 <i>d</i> (10.3)	5.84 <i>d</i> (10.3)	6.12 <i>d</i> (10.6)	6.16 <i>d</i> (10.5)	5.79 <i>d</i> (10.2)
3	—	—	—	—	—	—	—
4	—	—	—	—	—	—	—
5	2.37 <i>t</i> (5.1)	2.61 <i>m</i>	2.38 <i>m</i>	2.18 <i>m</i>	2.92 <i>m</i>	2.96 <i>m</i>	2.38 <i>m</i>
6	1.86 <i>m</i>	1.75 <i>m</i> 1.95 <i>m</i>	1.87 <i>m</i>	1.86 <i>m</i>	2.31 <i>m</i> 2.45 <i>m</i>	2.31 <i>m</i> 2.45 <i>m</i>	1.86 <i>m</i>
7	4.02 <i>bs</i>	5.26 <i>bs</i>	4.00 <i>bs</i>	5.26 <i>bs</i>	—	—	4.01 <i>bs</i>
8	—	—	—	—	—	—	—
9	2.43 <i>m</i>	2.39 <i>m</i>	2.39 <i>m</i>	2.36 <i>m</i>	2.96 <i>d</i> (6.6)	2.94 <i>d</i> (7.7)	2.38 <i>m</i>
10	—	—	—	—	—	—	—
11 α	1.59 <i>dd</i> (7.5, 9.2)	1.61 <i>m</i>	1.61 <i>m</i>	1.66 <i>m</i>	5.54 <i>dd</i> (6.6, 11.3)	5.51 <i>dd</i> (7.7, 10.3)	1.59 <i>dd</i> (14.2, 6.8)
11 β	2.42 <i>m</i>	2.51 <i>m</i>	2.44 <i>m</i>	2.46 <i>m</i>	—	—	2.55 <i>m</i>
12	4.95 <i>m</i>	4.93 <i>m</i>	4.96 <i>t</i> (8.0)	4.96 <i>m</i>	5.65 <i>d</i> (11.3)	5.66 <i>d</i> (10.3)	4.99 <i>m</i>
13	—	—	—	—	—	—	—
14	—	—	—	—	—	—	—
15	5.64 <i>bs</i>	5.43 <i>bs</i>	5.65 <i>d</i> (8.1)	5.49 <i>bs</i>	3.92 <i>s</i>	3.89 <i>s</i>	5.65 <i>bs</i>
16	2.58 <i>m</i>	2.49 <i>m</i>	2.57 <i>m</i>	2.47 <i>m</i>	1.85 <i>m</i> 2.47 <i>m</i>	2.10 <i>m</i> 2.38 <i>m</i>	2.48 <i>m</i>
17	3.00 <i>m</i>	2.92 <i>m</i>	2.98 <i>m</i>	2.94 <i>m</i>	2.99 <i>m</i>	2.99 <i>m</i>	3.05 <i>m</i>
18	1.01 <i>s</i>	1.01 <i>s</i>	1.03 <i>s</i>	1.02 <i>s</i>	0.95 <i>s</i>	0.97 <i>s</i>	1.00 <i>s</i>
19	1.16 <i>s</i>	1.19 <i>s</i>	1.15 <i>s</i>	1.18 <i>s</i>	0.96 <i>s</i>	0.94 <i>s</i>	1.16 <i>s</i>
20	—	—	—	—	—	—	—
21	—	—	6.04 <i>s</i>	6.05 <i>s</i>	5.81 <i>s</i>	—	—
22	6.95 <i>bs</i>	6.93 <i>bs</i>	5.97 <i>s</i>	5.95 <i>s</i>	5.88 <i>s</i>	6.84 <i>m</i>	6.87 <i>bs</i>
23	6.15 <i>bs</i>	6.14 <i>bs</i>	—	—	—	6.04 <i>m</i>	4.01 <i>bs</i>
28	1.14 <i>s</i>	1.04 <i>s</i>	1.13 <i>s</i>	1.19 <i>s</i>	1.06 <i>s</i>	0.97 <i>s</i>	1.14 <i>s</i>
29	1.06 <i>s</i>	1.04 <i>s</i>	1.05 <i>s</i>	1.05 <i>s</i>	1.07 <i>s</i>	1.07 <i>s</i>	1.06 <i>s</i>
30	1.16 <i>s</i>	1.21 <i>s</i>	1.15 <i>s</i>	1.21 <i>s</i>	5.34 <i>bs</i> 5.27 <i>bs</i>	5.31 <i>bs</i> 5.22 <i>bs</i>	1.15 <i>s</i>
Ac(CO)	—	—	—	—	—	—	—
Ac(CH ₃)	1.92 <i>s</i>	1.97 <i>s</i>	2.11 <i>s</i>	2.13 <i>s</i>	1.94 <i>s</i>	1.89 <i>s</i>	1.89 <i>s</i>
Ac(CO)	—	—	—	—	—	—	—
Ac(CH ₃)	—	1.95 <i>s</i>	—	1.97 <i>s</i>	1.91 <i>s</i>	1.85 <i>s</i>	—
OCH ₃	—	—	—	—	3.66 <i>s</i>	3.66 <i>s</i>	—
NH	—	—	—	—	—	—	6.36 <i>bs</i>

COSY spectrum. The $J_{9,11}$ and $J_{11,12}$ coupling constants of 6.6 and 11.3 Hz respectively indicated that the acetate groups were at C-11 β and C-12 α (Mulholland et al., 1998). This was confirmed by NOESY correlations. The side-chain was found to be the 21-hydroxy-23,21-butenolide ring as in **3** and **4**. This was confirmed by HMBC correlations between C-17 and H-21, C-21 and H-22 and C-22 and both H-17 β and H-21. Thus compound **5** was identified as 11-*epi*-21-hydroxytoonacilide. The 11-*epimer*, 21-hydroxytoonacilide has been isolated previously by Kraus and Grimmering (1980), and has been shown to exhibit feeding deterrents towards insects. The $J_{9,11}$ and $J_{11,12}$ coupling constants of 21-hydroxytoonacilide are both 4.1 Hz. (Kraus and Grimmering, 1980).

Compound **6**, C₃₁H₃₈O₁₁, was found to differ from **5** in having the 23-hydroxy-21,23-butenolide ring as in **1** and **2** and was identified as 11-*epi*-23-hydroxytoonacilide. The 11-*epimer* has been isolated by Kraus and Grimmering (1980), from *Toona ciliata* (Meliaceae).

Compound **7**, turrarparvin D, was shown by HRMS to have a molecular formula C₂₈H₃₇O₅N. The tetracyclic structure of **7** was found to be the same as turrarparvin A (**1**), with rings A–D intact, an α,β -unsaturated

ketone in ring A indicated by a pair of doublets at δ 7.00 (J = 10.2, H-1) and δ 5.79 (J = 10.2, H-2) and resonances at δ 157.49, δ 125.54 and δ 204.88 ascribed to C-1, C-2 and C-3 respectively. A Δ^{14} -double bond was indicated by resonances at δ 158.73 and δ 122.16 and a resonance ascribed to H-15 at δ 5.65. An acetate group was present at C-12 α and a hydroxy group at H-7 α , with H-12 β and H-7 β occurring at δ 4.99 and δ 4.01 respectively. This left a side chain of molecular formula C₄H₄NO, suggesting the presence of a lactam. The C-21 carbonyl resonance occurred at δ 175.74. The C-20 and C-22 alkene carbon resonances occurred at δ 138.81 and δ 139.80 respectively and the methylene carbon, C-23, occurred at δ 46.52. The COSY spectrum showed coupling between H-22 (δ 6.87, bs) and the 2H-23 resonance at δ 4.01(bs) and assignments were supported by the HMBC and NOESY spectra. It was initially not possible to see resonances due to C-21 and C-20 in the ¹³C NMR spectrum, but after lengthy re-running of the spectrum (120 h) and with the help of correlations seen in the HMBC spectrum, these peaks could be found. There has been one previous report of salannolactam-(21) a related lactam, by Kraus et al., from *Azadirachta indica* (Kraus et al., 1987). This lactam also co-occurred

Table 2

¹³C NMR spectral data for compounds **1–7** (100 MHz, CDCl₃) (values in parentheses are due to the minor epimer)

Carbon	1	2	3	4	5	6	7
1	157.45 (157.41)	157.39	157.11	156.94	151.96 (152.07)	152.14 (152.00)	157.49
2	125.60	125.53	125.76 (125.79)	125.75	125.87	125.97 (125.88)	125.54
3	204.09	204.88	204.83	204.30	204.30 (204.19)	204.09	204.86
4	40.09	39.85	39.96	44.05	46.24	46.19	40.09
5	44.36	46.00	44.35	46.05	44.91	45.12	44.64
6	24.28	23.63	24.46	23.71	31.24	31.28	24.23
7	71.22 (71.21)	73.78	71.49 (71.26)	73.93 (73.77)	174.14	174.22	71.20
8	44.73	42.56	44.83	42.70	135.68 (135.49)	136.17	44.36
9	38.42 (38.44)	39.70	38.03	39.38	52.53 (52.65)	52.82	38.47
10	44.14	44.03	44.15	39.71	42.04	42.01	44.13
11	25.39	25.56	25.62	25.86	70.43 (71.15)	71.40	25.40
12	76.94 (76.91)	77.17	77.61	77.76	75.32 (75.71)	75.09 (75.02)	76.95
13	51.31 (51.47)	51.22 (51.35)	52.43 (52.39)	52.32	45.91	46.09 (45.83)	50.88
14	158.46 (158.80)	156.08 (156.38)	157.93	155.57	71.30 (70.61)	71.08	158.73
15	121.75 (121.95)	121.08 (120.91)	122.17 (121.75)	121.46	59.47 (58.97)	59.83 (59.52)	122.16
16	35.65 (35.35)	35.62	35.54 (35.91)	35.42	32.70 (32.30)	31.28 (29.84)	35.67
17	48.80 (48.62)	48.87	50.69 (51.15)	51.13 (50.77)	40.48 (40.18)	38.92 (38.19)	48.82
18	15.15 (15.23)	15.02 (15.08)	15.94	15.92	13.02	13.41 (13.45)	15.34
19	18.83	18.87 (18.85)	18.85	18.89 (18.99)	20.95	21.30	18.80
20	137.03 (136.94)	136.99	168.56 (167.48)	168.86	166.92 (167.29)	135.04 (135.64)	138.81
21	171.38 (171.33)	171.67	99.16 (99.55)	99.21 (99.63)	99.30 (99.37)	170.16 (170.73)	175.74
22	144.92 (145.57)	145.06 (145.74)	120.52 (118.26)	120.39 (121.23)	118.32 (120.35)	147.69 (146.57)	139.80
23	96.40 (96.12)	96.20 (96.58)	170.60 (170.72)	170.61	170.40	95.65 (96.08)	46.52
28	26.94	26.82	26.99	26.87	22.90 (22.87)	22.96	26.95
29	21.45	21.13	21.42	21.35	22.77	22.78	21.09
30	27.82 (27.86)	27.60	28.07	27.79 (27.65)	122.29 (121.90)	121.36	27.75
Ac(CO)	170.69 (170.72)	170.90	171.82	170.45	169.62	169.85	170.64
Ac(CO)	—	170.03	—	169.92	20.56	20.65	—
Ac(CH ₃)	21.05	21.25	21.42	21.24	170.19 (171.55)	169.97	21.45
Ac(CH ₃)	—	21.19	—	21.13	21.32	20.56	—
OCH ₃	—	—	—	—	52.21	52.17	—

with compounds with hydroxybutenolide side-chains as in this case. The chemical shifts of C-20 to C-23 in salannolactam occurred at δ 140.9, δ 175.2, δ 138.0 and δ 46.8 and are in reasonably close agreement with chemical shifts for the same carbons in turrarparvin D (δ 138.8, δ 175.7, δ 139.8 and δ 46.5).

3. Experimental

Seeds of *Turraea parvifolia* Deft. (Meliaceae) were collected from Chemolingot, Kenya identified by Mr. John Kisang and a voucher specimen retained (PC1) at Egerton University, Kenya. The ground seed (118.31 g) was left in methanol for 4 days. The methanol extract (9.27 g) yielded, after column chromatography over silica gel (Merck 9385) using 10% ethyl acetate in dichloromethane, turrarparvin A (12 α -acetoxy-7 α ,23-dihydroxy-24,25,26,27-tetranor-3-oxoapotirucalla-1,14,20(22)-trien-21,23-olide), **1**, (12.2 mg), 12 α -acetoxyazadironolide, **2**, (13.2 mg), turrarparvin B (12 α -acetoxy-7 α ,21-dihydroxy-24,25,26,27-tetranor-3-oxoapotirucalla-1,14,20(22)-trien-23,21-olide), **3**, (8.9 mg), turrarparvin C (7 α ,12 α -diacetoxy-21-hydroxy-24,25,26,27-tetranor-3-oxoapotirucalla-1,14,20(22)-trien-23,21-olide), **4**, (9.3 mg), 11-*epi*-21-hydroxytoonacilide, **5**, (10.3 mg), 11-*epi*-23-hydroxytoonacilide, **6**, (13.7 mg) and turrarparvin D (12 α -acetoxy-7 α ,hydroxy-24,25,26,27-tetranor-3-oxoapotirucalla-1,14,20(22)-trien-21,23-lactam), **7**, (5.4 mg). NMR data for compounds **1–7** is given in [Tables 1 and 2](#).

IR spectra were recorded with a Nicolet Impact 400 D spectrometer on sodium chloride plates and calibrated against an air background. HRMS were obtained using a Kratos High Resolution MS 9/50 spectrometer at the Cape Technikon. ^1H and ^{13}C NMR spectra were recorded on a Varian Unity Inova 400 MHz NMR spectrometer. Optical rotations were measured using a Perkin Elmer 241 Polarimeter with tube 10 cm in length. Melting points (uncorr.) were determined using a K  fler hot stage apparatus.

3.1. Turrarparvin A (12.2 mg), **1**

White crystalline; mp 108–110 °C; HRMS (rel. int.) m/z : at 466.2355 $[\text{M}-\text{H}_2\text{O}]^+$ ($\text{C}_{28}\text{H}_{34}\text{O}_6$ req. 466.2367, 2), 424.2251 $[\text{M}-\text{CH}_3\text{COOH}]^+$ (26), 406.2119 $[\text{M}-\text{H}_2\text{O}-\text{CH}_3\text{COOH}]^+$ (20), 391 (33), 373 (13), 345 (6), 295 (8), 269 (19), 239 (3), 211 (22), 150 (94), 109 (34), 71 (55) and 57 (100); IR: $\nu_{\text{max}}(\text{NaCl}) \text{ cm}^{-1}$: 3425, 2923, 2856, 1736, 1658 1462, 1250, 1038; $[\alpha]_{\text{D}}^{23} + 36.8^\circ$ (CHCl_3 ; c 0.20).

3.2. 12 α -Acetoxyazadironolide (13.2 mg), **2**

White crystalline; mp 97–99 °C; HRMS (rel. int.) m/z : 526.2560 $[\text{M}]^+$ ($\text{C}_{30}\text{H}_{38}\text{O}_8$ req. 526.2567, 1), 508.2510

$[\text{M}-\text{H}_2\text{O}]^+$ (1), 466.2366 $[\text{M}-\text{CH}_3\text{COOH}]^+$ (9), 406.2138 $[\text{M}-\text{CH}_3\text{COOH}-\text{CH}_3\text{COOH}]^+$ (14), 391 (15), 373 (7), 345 (3), 269 (14), 257 (28), 239 (31), 211 (17), 150 (100), 109 (17) and 69 (4); IR: $\nu_{\text{max}}(\text{NaCl}) \text{ cm}^{-1}$: 2971, 2860, 1732, 1474, 1393, 1252, 1135, 1034; $[\alpha]_{\text{D}}^{23} + 63.1^\circ$ (CHCl_3 ; c 0.20).

3.3. Turrarparvin B (8.9 mg), **3**

White crystalline; mp 142–144 °C; HRMS (rel. int.) m/z : 484.2449 $[\text{M}]^+$ ($\text{C}_{28}\text{H}_{36}\text{O}_7$ req. 484.2461, 3), 466.2346 $[\text{M}-\text{H}_2\text{O}]^+$ (24), 424.2240 $[\text{M}-\text{CH}_3\text{COOH}]^+$ (66), 388.2023 $[\text{M}-\text{H}_2\text{O}-\text{H}_2\text{O}-\text{CH}_3\text{COOH}]^+$ (21), 373 (15), 357 (36), 337 (18), 315 (12), 257 (19), 239 (33), 219 (66), 179 (78), 150 (60), 137 (99), and 109 (100); IR: $\nu_{\text{max}}(\text{NaCl}) \text{ cm}^{-1}$: 3420, 2926, 2859, 1742, 1670, 1465, 1254, 1115, 1042; $[\alpha]_{\text{D}}^{23} + 29.1^\circ$ (CHCl_3 ; c 0.09).

3.4. Turrarparvin C (9.3 mg), **4**

White crystalline; mp 117–119 °C; HRMS (rel. int.) m/z : 526.2573 $[\text{M}]^+$ ($\text{C}_{30}\text{H}_{38}\text{O}_8$ req. 526.2567, 5), 508.2492 $[\text{M}-\text{H}_2\text{O}]^+$ (20), 466.2361 $[\text{M}-\text{CH}_3\text{COOH}]^+$ (47), 406.2147 $[\text{M}-\text{CH}_3\text{COOH}-\text{CH}_3\text{COOH}]^+$ (44), 388.2030 $[\text{M}-\text{H}_2\text{O}-\text{CH}_3\text{COOH}-\text{CH}_3\text{COOH}]^+$ (42), 373 (20), 337 (13), 279 (14), 239 (60), 238 (37), 211 (19), 150 (80), 137 (90) and 57 (100); IR: $\nu_{\text{max}}(\text{NaCl}) \text{ cm}^{-1}$: 3388, 2915, 2849, 1741, 1670, 1389, 1246, 1130, 1042; $[\alpha]_{\text{D}}^{23} + 35.2^\circ$ (CHCl_3 ; c 0.14).

3.5. 11-*epi*-21-Hydroxytoonacilide (10.3 mg), **5**

White crystalline; mp 124–126 °C; HRMS (rel. int.) m/z : 586.2416 $[\text{M}]^+$ ($\text{C}_{31}\text{H}_{38}\text{O}_{11}$ req. 586.2414, 97), 526.2204 $[\text{M}-\text{CH}_3\text{COOH}]^+$ (22), 508.2097 $[\text{M}-\text{H}_2\text{O}-\text{CH}_3\text{COOH}]^+$ (11), 466.1991 $[\text{M}-2\times\text{CH}_3\text{COOH}]^+$ (19), 484 (8), 467 (27), 449 (27), 391 (41), 297 (54), 281 (23), 239 (36), 210 (42), 149 (100), 105 (86) and 69 (61); IR: $\nu_{\text{max}}(\text{NaCl}) \text{ cm}^{-1}$: 2928, 2849, 1753, 1674, 1383, 1240, 1048; $[\alpha]_{\text{D}}^{23} + 26.0^\circ$ (CHCl_3 ; c 0.10).

3.6. 11-*epi*-23-Hydroxytoonacilide (13.7 mg), **6**

White crystalline; mp 139–142 °C; HRMS (rel. int.) m/z : 586.2419 $[\text{M}]^+$ ($\text{C}_{31}\text{H}_{38}\text{O}_{11}$ req. 586.2414, 61), 568.2309 $[\text{M}-\text{H}_2\text{O}]^+$ (14), 526.2203 $[\text{M}-\text{CH}_3\text{COOH}]^+$ (14), 466.1991 $[\text{M}-2\times\text{CH}_3\text{COOH}]^+$ (24), 449 (25), 433 (20), 391 (54), 311 (43), 297 (100), 257 (25), 239 (46), 149 (84), 137 (57), 105 (68) and 57 (85); IR: $\nu_{\text{max}}(\text{NaCl}) \text{ cm}^{-1}$: 2929, 2852, 1753, 1670, 1382, 1255, 1056; $[\alpha]_{\text{D}}^{23} + 33.8^\circ$ (CHCl_3 ; c 0.07).

3.7. Turrarparvin D (5.4 mg), **7**

White crystalline, mp 131–133 °C, HRMS (rel. int.) m/z : 467.2662 $[\text{M}]^+$ ($\text{C}_{28}\text{H}_{37}\text{NO}_5$ req. 467.2672, 1),

449.2566 $[M-H_2O]^+$ (2), 407.2460 $[M-CH_3COOH]^+$ (41), 421 (10), 406 (16), 392 (80), 374 (74), 313 (7), 252 (28), 227 (100), 188 (22), 137 (61), 135 (29) and 82 (69); IR: $\nu_{max}(NaCl)$ cm^{-1} : 2923, 2856, 1730, 1249, 1137; $[\alpha]_D^{23} + 29.8^\circ$ ($CHCl_3$; c 0.08).

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References

- Adul, G.O., Bentley, M.D., Benson, B.W., Huang, F., Gelbaum, L., Hassanali, A., 1993. Two new pterianin-class limonoids from *Turraea mombasana*. *J. Nat. Prod.* 56, 1414–1417.
- Akinniyi, J.A., Connolly, J.D., Mulholland, D.A., Rycroft, D.S., Taylor, D.A.H., 1986. Limonoids extractives from *Turraea floribunda* and *T. obtusifolia*. *Phytochemistry* 25, 2187–2189.
- Bentley, M.D., Adul, G.O., Alford, A.R., Huang, F., Gelbaum, L., Hassanali, A., 1995. An insect antifeedant limonoid from *Turraea nilotica*. *J. Nat. Prod.* 58, 748–750.
- Bentley, M.D., Gaul, F., Rajab, M.S., Hassanali, A., 1992. Tetratriterpenes from *Turraea robusta*. *J. Nat. Prod.* 55, 84–87.
- Blundell, M., 1987. *Wild Flowers of East Africa*. William Collins Sons & Co. Ltd, London.
- Chiplunkar, Y.G., Nagasampagi, B.A., Tavale, S.S., Puranik, V.G., 1993. Villostrol, 3 β ,5 β -dihydroxy-20-pregnen-6-one, steroid from *Turraea villosa*. *Phytochemistry* 33, 901–903.
- Fraser, L.N., Mulholland, D.A., Nair, J.J., 1994. Limonoids from the seed of *Turraea floribunda*. *Phytochemistry* 35, 455–458.
- Fraser, L.N., Mulholland, D.A., Taylor, D.A.H., 1995. The chemotaxonomic significance of the limonoids, nymania-1, in *Turraea obtusifolia*. *S. Afr. J. Bot.* 61, 281–282.
- Kraus, W., Grimmer, W., 1980. 23-(*R,S*)-Hydroxytoonacilid und 21-(*R,S*)-hydroxytoonacilid, zwei neue B-seco-tetratriterpenoide mit insektenfrasshemmender Wirkung aus *Toona ciliata* M. J. Roem. *Var. Australis* (Meliaceae). *Nouv. J. Chim.* 4, 651–655.
- Kraus, W., Klenk, A., Bokel, M., Vogler, B., 1987. Tetratriterpenoid-Lactame mit insektenfrasshemmender Wirkung aus *Azadirachta indica* A. Juss (Meliaceae). *Liebigs Ann. Chem.* 337–340.
- Mulholland, D.A., Monkhe, T.V., 1993. Two glabretal-type triterpenoids from the heartwood of *Aglaia ferruginea*. *Phytochemistry* 34, 579–580.
- Mulholland, D.A., Monkhe, T.V., Coombes, P.H., Rajab, M.S., 1998. Limonoids from *Turraea holstii* and *Turraea floribunda*. *Phytochemistry* 49, 2585–2590.
- Mulholland, D.A., Monkhe, T.V., Taylor, D.A.H., Rajab, M.S., 1999. Triterpenoids from *Turraea holstii*. *Phytochemistry* 52, 123–126.
- Mulholland, D.A., Taylor, D.A.H., 1988. Protolimonoids from *Turraea nilotica*. *Phytochemistry* 27, 1220–1221.
- Rajab, M.S., Bentley, M.D., Hassanali, A., Chapya, A., 1988. A new limonoid from *Turraea robusta*. *Phytochemistry* 27, 2353–2355.
- Siddiqui, B.S., Faizi, G.S., Rasheed, M., 1999. Triterpenoids of the fruit coats of *Azadirachta indica*. *J. Nat. Prod.* 62, 1006–1009.
- Styles, B.T., White, F., 1991. Meliaceae. In: Polhill, R.M. (Ed.), *Flora of Tropical East Africa*. Royal Botanic Gardens/KEW. A.A. Balkema, Rotterdam, p. 14.
- Torto, B., Bentley, M.D., Cole, B.J.W., Hassanali, A., Huang, F., Gelbaum, L., Vanderveer, D.G., 1995. Limonoids from *Turraea floribunda*. *Phytochemistry* 40, 239–243.
- Torto, B., Hassanali, A., Nyandat, E., Bentley, M.D., 1996. A limonoid from *Turraea floribunda*. *Phytochemistry* 42, 1235–1237.